


REMARKS

The specification has been amended to incorporate the sequence identifiers pursuant to 37 C.F.R. § 1.821(d) and the sequence listing pursuant to 37 C.F.R. § 1.821(c). The above-made amendments do not introduce new matter.

Applicants respectfully submit that this Amendment be made of record in the file history of the instant application.

Respectfully submitted,

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VERSION WITH MARKINGS TO SHOW CHANGES MADE

On page 14, lines 28 and 29, please amend the paragraph as follows:

Fig. 4 shows sequences of the utilized regions i.e. A', B and C: HCV (SEQ ID NO: 6) and Human (SEQ ID NO: 7).

On page 15, lines 4-11, please amend the paragraph as follows:

Fig. 7 shows a particularly suitable region (SEQ ID NO: 8) of the HCV genome for performing the method according to the invention and a sequence from which the primer and probe sequences are preferably selected. This second sequence is taken from the non-human pathogenic virus HGBV-B (SEQ ID NO: 9). The selected primer and probe sequences are therefore sequences that are not specific for HCV (M. Med. Virol. 48, 60-67).

On page 15, lines 12 and 13, please amend the paragraph as follows:

Fig. 8 to 10 show preferred sequences for primers and probes for the HCV test: MPF1 (SEQ ID NO: 10), MPF1+1 (SEQ ID NO: 11), MPF2 (SEQ ID NO: 12), HCV_1A (SEQ ID NO: 13), MPR1_rev&compl (SEQ ID NO: 14), MPR2_rev & compl (SEQ ID NO: 15), HCVMCR02_rev&compl (SEQ ID NO: 16), Foreward primer CK10/Reverse primer CK20 (SEQ ID NOs: 17 and 18), Foreward primer CK11/Reverse primer CK20 (SEQ ID NOs: 19 and 20), Foreward primer CK10-1/Reverse primer CK20-1 (SEQ ID NOs: 21 and 22), Foreward primer CK11-1 (SEQ ID NO: 23), Foreward primer CK 10-2/Reverse primer CK20-2 (SEQ ID NOs: 24 and 25), Foreward primer CK11-2 (SEQ ID NO: 26), Reverse primer CK21 (SEQ ID NO: 27), Foreward primer CK10-1/Reverse Primer CK21-1 (SEQ ID NOs: 28 and 29), Foreward primer CK11-1 (SEQ ID NO: 30), Reverse primer CK21-2 (SEQ ID NO: 31), Reverse primer CK21-3 (SEQ ID NO: 32), Foreward primer CK12/Reverse primer CK22 (SEQ ID NOs: 33

and 34), Foreward primer CK12-1/Reverse primer CK22-1 (SEQ ID NOs: 35 and 36), Reverse primer CK22-2 (SEQ ID NO: 37), Reverse Primer CK22-3 (SEQ ID NO: 38), Foreward primer CK12-2/Reverse primer CK22-4 (SEQ ID NOs: 39 and 40), Reverse primer CK22-5 (SEQ ID NO: 41), Reverse primer CK23 (SEQ ID NO: 42), Reverse primer CK23-1 (SEQ ID NO: 43), Reverse primer CK23-2 (SEQ ID NO: 44), Reverse primer CK23-3 (SEQ ID NO: 45), Reverse primer CK24 (SEQ ID NO: 46), Reverse primer CK24-1 (SEQ ID NO: 47), Reverse primer CK24-2 (SEQ ID NO: 48), Reverse primer CK24-3 (SEQ ID NO: 49), HCV (SEQ ID NO: 93) and HGBV-B (SEQ ID NO: 94).

On page 55, lines 10-13, please amend the paragraph as follows:

Two different ruthenium-labelled probes were used for the hybridization:

PNA-probe: Ru-(Ser)₂-TCCAGGACCC-Ser-Gly (SEQ ID NO: 92)

DNA-probe: 5'-Ru-CTCCAGGACCCC-3', (SEQ.ID.NO.5)

On page 63, please amend the table as follows:

primer	sequence	position	amplicon
SK 462 (SEQ ID NO: 50)	5'-AGTTGGAGGACATCAAGCAGCCATGCAAAT-3'	1359-1388 (30)	142 bp
SK 431 (SEQ ID NO: 51) (gag)	5'-TGCTATGTCAGTTCCCCTTGGTTCTCT-3'	1474-1500 (27)	
SK 102 (SEQ ID NO: 52)	5'-ATCAATGAGGAAGCTGCAGA-3'	1402-1421 (20)	
RAR 1032 (SEQ ID NO: 53)	5'-GAGACACCAGGAATTAGATATCAGTACAATGT-3'	2961-2992 (32)	169 bp
RAR 1033 (SEQ ID NO: 54) (pol)	5'-CTAAATCAGATCCTACATATAAGTCATCCATGT-3'	3097-3129 (33)	
RAR 1034 (SEQ ID NO: 55)	5'-CCACAAGGATGGAAGGATCACCAGCTATATTCCA-3'	2997-3031 (35)	
GH A1F (SEQ ID NO: 56)	5'-TGTACCAGTAAAATTAAAGCCAG	2570-2592 (23)	

GH A1R (SEQ ID NO: 57) (pol)	5'-GGCCATTGTTTAACTTTTGG	2604-2623 (20)	54 bp
GH A1P (SEQ ID NO: 58)	5'-AGGAATGGATGGC	2591-2603 (13)	
GH A2F (SEQ ID NO: 59)	5'-TACCTGGCATGGGTACCAGC	4143-4162 (20)	63 bp
GH A2R (SEQ ID NO: 60) (pol)	5'-GACTAATTTATCTACTTGTTTCATTTTC	4180-4205 (26)	
GH A2P (SEQ ID NO: 61)	5'-CACACAAAGGAATTGGAG	4162-4179 (18)	
GH A3F (SEQ ID NO: 62)	5'-TTTGGAATTCCTACAATCC	4644-4663 (20)	59 bp
GH A3R (SEQ ID NO: 63) (pol)	5'-AATTCTTTATTCATAGATTCTACTAC	4677-4702 (26)	
GH A3P (SEQ ID NO: 64)	5'-CCCAAAGTCAAGGAG	4663-4677 (15)	
GH A4F (SEQ ID NO: 65)	5'-TCAAAATTTTCGGGTTTATTACAG	4889-4912 (24)	63 bp
GH A4R (SEQ ID NO: 66) (pol)	5'-AGCTTTGCTGGTCCTTTCCA	4932-4951 (20)	
GH A4P (SEQ ID NO: 67)	5'-GGACAGCAGAAATCCACTT	4913-4931 (19)	
GH A5F (SEQ ID NO: 68)	5'-GGAAAAGGTCTATCTGGCATGGGT	4133-4156 (24)	72 bp
GH A5R (SEQ ID NO: 69) (pol)	5'-ACTAATTTATCTACTTGTTTCATTTCTC	4177-4204 (28)	
GH A5P (SEQ ID NO: 70)	5'-ACCAGCACACAAAGGAATTG	4157-4176 (20)	
GH A6F (SEQ ID NO: 71)	5'-GCAACTAGATTGTACACATTTAGAAG	4412-4437 (26)	74 bp
GH A6R (SEQ ID NO: 72) (pol)	5'-CTTCTATATATCCACTGGCTACATG	4461-4485 (25)	
GH A6P (SEQ ID NO: 73)	5'-GAAAAGTTATCCTGGTAGCAGTT	4438-4460 (23)	

On page 65, please amend the table as follows :

	primer/probe	sequence	position	amplicon length
Ref	HBV-Forward (SEQ ID NO: 74) HBV-Reverse (SEQ ID NO: 75) capture probe (SEQ ID NO: 76)	5'-GGAGTGTGGATTCGCACT-3' 5'-TGAGATCTTCTGCGACGC-3' 5'-AGACCACCAAATGCCCCTAT-3'	2267-2284 (18) 2419-2436 (18) 2297-2316 (20)	170 bp
1	GHBV-1F (SEQ ID NO: 77) GHBV-1R (SEQ ID NO: 78) capture probe 1P (SEQ ID NO: 79)	5'-CCACCAAATGCCCCTAT-3' 5'-CCCGTCGTCTAACAACAG-3' 5'-CTTATCAACACTTCCGGAACTA-3'	2300-2316 (17) 2340-2357 (18) 2317-2339 (23)	58 bp
2	GHBV-2F (SEQ ID NO: 80) GHBV-2R (SEQ ID NO: 81) capture probe 2P (SEQ ID NO: 82)	5'-GCGGGGTTTTTCTTGTT-3' 5'-TCTAGACTCTGCGGTATTGTG-3' 5'-TTGACAAGAATCCTCA-3'	203-219 (17) 232-252 (21) 218-233 (16)	50 bp
3	GHBV-3F (SEQ ID NO: 83) GHBV-3R (SEQ ID NO: 84) capture probe 3P (SEQ ID NO: 85)	5'-GATCCCCAACCTCCAATC-3' 5'-CAGCGATAACCAGGACAAAT-3' 5'-ACTCACCAACCTCCTGTCCTCCA-3'	315-332 (18) 356-375 (20) 333-355 (23)	61 bp
4	GHBV-4F (SEQ ID NO: 86) GHBV-4R (SEQ ID NO: 87) capture probe 4P (SEQ ID NO: 88)	5'-ACTTCTTTCTTCCGTCAGA-3' 5'-AAGGCTTCCCGATACAGAG-3' 5'-GATCTCCTAGACACCGCCTCGG-3'	1965-1984 (20) 2007-2015 (19) 1985-2006 (22)	61 bp
5	GHBV-5F (SEQ ID NO: 89) GHBV-5R (SEQ ID NO: 90) capture probe 5P (SEQ ID NO: 91)	5'-CAGCCAACCAGGTAGGAGTG-3' 5'-CCGTGTGGAGGGGTGAAC-3' 5'-GGAGCATTCGGGCCAGG-3'	3014-3033 (20) 3051-3068 (18) 3034-3050 (17)	55 bp